

Aesculus indica: An ethnomedicinal, phytochemical, and pharmacological review

Som Kailash Madhvi^{1*}, Javaid Iqbal¹ and Mohd Younis²

¹Department of Zoology and Applied Aquaculture, Barkatullah University, Bhopal 462026, Madhya Pradesh, India

²Department of Human Genetics and Molecular Biology, Bharathiar University, Coimbatore 641046, Tamil Nadu, India

Received 27 March 2021; revised received 06 May 2023; accepted 18 May 2023

Aesculus indica, commonly known as Indian horse chestnut, is an indigenous tree of the family Sapindaceae, distributed in Nepal, India, Pakistan, and Afghanistan. As an ethnomedicine, its seeds are used to cure jaundice, chest diseases, stomach and joint pain, roots for leucorrhea, and bark for fever and anaemia. The phytochemicals of class phenolic, triterpenoid, glycoside, saponin, and fatty acid have been identified from their different parts and various biological activities such as antioxidant, anti-diabetic, hypocholesterolemic, hypoglycaemic, anti-nociceptive, anti-inflammatory, neuro-depressive, spasmolytic, immunomodulatory, anticancer, enzyme inhibitory, antimicrobial and insecticidal have been reported, which reflected its medicinal importance. However, most biological activities are on the extracts or fractions and need more in-depth study. Therefore, further research is required on the isolation, identification, and characterization of active phytochemicals from the extracts with their mechanism of pharmacological action and detailed toxicological evaluation. In this review, we aim to provide all information regarding traditional uses, morphology, phytochemistry, pharmacology, and toxicity of *A. indica* and address the research gaps; therefore, will provide a source of information to the researchers for further studies.

Keywords: *Aesculus indica*, Ethnomedicinal, Pharmacology, Phytochemicals, Sapindaceae

IPC code; Int. cl. (2021.01)- A61K 36/00, A61K 36/77, A61P

Introduction

Aesculus indica is an indigenous tree of woodland habitat usually known as Indian horse chestnut or Himalayan chestnut¹. In different regions, it is called by various names, such as *hanudon* in Kashmir, *goon* in Himachal Pradesh, *khajushing* in Bhutan, *pangro* in Nepal, and *jawaz* in Pakistan²⁻⁴. This plant belongs to the genus *Aesculus* which contains 12 species distributed in America, Europe, and Asia⁵. *A. indica* has many folk medicinal uses such as indigestion, stomach pain, joint pain, venereal, chest diseases, skin ulcer, haemorrhoids, and jaundice⁶⁻⁹. The seed extract of *A. indica* was effective against human nasopharynx epidermoid carcinoma, P-388 lymphocytic leukaemia, and has antimicrobial and haemagglutination properties¹⁰⁻¹². The seed oil has anti-inflammatory properties¹³. The seed shell adsorbs cadmium (II) ions from an aqueous solution¹⁴ and leaves have antioxidant properties¹⁵. The preliminary phytochemical screening of *A. indica* showed the presence of phenol, flavonoid, alkaloid¹⁶, coumarin, protein, amino acid, steroid, carbohydrate, saponin, tannin, terpenoids in the leaf extracts¹⁷; glycoside,

saponin, flavonoid, tannin¹⁸ carbohydrate and phenol in the seed extracts¹⁹. The current study aims to review folk uses, phytochemistry, and pharmacology of *A. indica*.

Methodology

Literature was searched using plant names and synonyms in Pubmed, Google Scholar, and publisher databases such as Bentham, Springer, and ScienceDirect. Books, conference papers, patents, and a few websites such as Kew botanical garden, Centre for Agriculture and Bioscience International, India Biodiversity Portal, Tropicos, Pubchem and ChemSpider were also utilized.

Plant description

Geographical distribution

A. indica is distributed in Asian countries along the northwest Himalayas from west Nepal, north India, and northwest Pakistan to northeast Afghanistan at an altitude of 3000 m²⁰. It is distributed in India's moist and shady valleys of Kashmir, Uttarakhand, and Himachal Pradesh. In Himachal Pradesh, it is found in the forests of districts Sirmour, Mandi, Shimla, Chamba, Solan, Kangra, and Kullu²¹.

*Correspondent author
Email: somkailash86@gmail.com

Taxonomy

Kingdom: Plantae, phylum: tracheophyta, class: magnoliopsida, order: sapindales, family: sapindaceae, genus: *Aesculus*, species: *Aesculus indica*²². Synonyms: *Aesculus indica* var. *concolor* Browicz, *Pavia indica* Wall. ex Cambess²³.

Morphology

A. indica (Fig. 1a-g) is a medium to large-sized tree with a straight, cylindrical trunk and a round to broadly spreading canopy. It has a 25 - 30 m average height and a 2.5 m diameter trunk. The bark is smooth grey and peels off from upwards in narrow bands when it ages. The petiole is about 8 - 19 cm long, having 5 - 9 leaflets; leaves are 15 - 25 cm long and up to 8 cm wide, opposite, digitate, exstipulate, glabrous, serrated, oblong to lanceolate and narrowed at the base. The calyx is gamosepalous, tubular, 5 - lobed, 4 - 8 mm long, with short and rounded lobes. Flowers are white-yellowish streaked coloured, having 3.5 mm length and 2 - 2.5 cm diameter, pedicellate, perigynous, bisexual, and present in about 32 cm long compound raceme inflorescence average of 385 flowers. The flower has four unequal petals: the upper petals are (oblanceolate) 15 - 24 mm long, and the lateral petals (oblong to obovate) 15 - 20 mm long. Ovary with a single style, androecium is polyandrous (5 - 9 stamens, 3.5 cm long), perigynous,

glabrous and up to 30 mm long. The gynoecium is tomentose, fruit capsule is ovoid, about 2 cm long and 4.1 cm in diameter, with rusty brown spots on the epicarp. The capsule contains 1 - 2 shiny dark brown coloured with white scars seeds and two lemon white cotyledons. Flowering occurs in April-May, and seeds mature in October²⁴⁻²⁹.

Traditional uses

A. indica is used for several purposes, its seeds are eaten by wild animals and humans³⁰ but are considered harmful for humans if consumed without any previous treatment³¹. Pre-treatments include water soaking, blanching, and cooking, which removes anti-nutritional factors like saponins³². In Himachal Pradesh, seeds are crushed and treated with water to remove bitter contents (saponins), dried, and prepared into flour/powder called *tatwakhar*, and various food items like porridge, *chappatis* are prepared from it and sold in the markets³³. Seeds contain saponins, so they are used as an alternative soap³⁴. The leaves are fed to livestock and collected for the winter when there is a scarcity of green fodder³⁵. *A. indica* is also used as a fish toxicant³⁶. Its flowers are used in apiculture as bee forage³⁷, and wood is used in the fences³⁸, as fuel³⁹, and in the preparation of various household articles and agricultural tools like hoe and yoke⁴⁰. Despite the above uses *A. indica* is also



Fig. 1 — *Aesculus indica*, a) Whole plant; b) Inflorescence; c) Stem bark; d) Unripe fruit; e) Ripe fruit; f) Leaflets; and g) leaf.

utilized to cure different types of health problems as leaves extract are used to cure whooping and cough⁴¹, fruits are used to kill stomach worms⁴², prevent hair fall⁴³, paste applied to cure mumps⁴⁴, and its powder is overcome general weakness⁴⁵, sometimes powder is mixed with husk and fed to animals to get relief from abdominal pain⁴⁶. The seed powder roasted in butter is used to prevent excess bleeding and pain in menses, also given to women during the pre- and post-period of pregnancy⁴⁷. The seed paste relieves rheumatic pain⁴⁸ and seed oil is applied to relieve headaches and skin infections⁴⁹. The bark is used to treat hepatitis⁵⁰, anaemia, chest pain⁵¹, dislocated joints⁵², and curing fever⁵³. The roots are used for the treatment of leucorrhoea⁵⁴.

Physicochemical composition

A. indica has several traditional uses, as discussed in the previous section. One of those is that the seeds are washed with water to remove saponins before using as food. So the researcher studied and reported various methods (Soxhlet, percolation, boiling, roasting, microwave, and pressure cooking) to optimize toxic/bitter contents and nutritional mineral composition⁵⁵⁻⁵⁷. Also, some quality evaluation studies have been carried out on the products of seed flour^{58,59} and results are found to be almost similar to standard recipes⁶⁰. *A. indica* contains various nutrients and minerals^{61,62} (Table 1) such as phosphorus, potassium⁵⁶, calcium, manganese, zinc, iron, copper^{56,57}, protein, carbohydrates and fat^{63,64}.

Phytochemistry

Many studies carried out on different parts of *A. indica* have confirmed the presence of compounds that belongs to the category of phenol, flavonoid, flavonol glycoside, terpenoid, terpenoid glycoside, fatty acid and phytosterol.

Phenolic compounds

The presence of chlorogenic acid, phloroglucinol, hydroxybenzoic acid was detected in various (fruit) fractions and isolated a phenolic, mandelic acid; a flavonol, quercetin from the ethyl acetate fraction⁶⁵ (Fig. 2a-d) and flavonol glycosides, astragalgin⁶⁶, rutin⁶⁷ and quercitrin from the chloroform leaf extract⁶⁸ (Fig. 2g-i). A phytosterol, β -sitosterol (Fig. 2j) was also isolated from the hexane leaves extract⁶⁸. Two other phenols, catechol and epicatechin are also reported^{52,69} (Fig. 2e,f).

Triterpenoids

The triterpenoids, barringtogenol-C, proto-aescigenin, aescigenin (Fig. 3a-c) and acyl-triterpenoids, 21-angeloylbarringtogenol-C, 21, 22 diangeyleylbarringtogenol-C and 22-angeloyl-R1-barrigenol are characterized from the hydrolysate mixture of the seeds of *A. indica*^{5,70} (Fig. 3d,e). The triterpenoid glycosides, aesculusides A⁷¹, aesculusides B⁷² were isolated from the methanol seed extract and 3-O-[[Beta-3-D-glucopyranosyl(1-2)]]{Beta-D-glucopyranosyl (1-4)}-Beta-D-gluconopyranosyl] 21, 22-diangeloyl-barringtogenol C was isolated from the 90% methanol seed-coats extract⁷³ (Fig. 3f-h). A triterpene saponin aescin was isolated from the chloroform leaves extract⁶⁸, aqueous ethanol seed extract⁷⁴, and an isomer β -aescin from hydroalcoholic seed extract⁷⁵. The chemical structures are depicted in Fig. 3^{5,73,76}.

Fatty acid

The presence of saturated long-chain fatty acids, palmitic acid, myristic acid, stearic acid, arachidic acid, lauric acid, and oleic acid (Fig. 4a-f) was detected in the gas chromatography of the saponified hexane extracts of the seeds, leaves, and husks⁷⁷. In another study, palmitoleic acid, 10-heptadecenoic acid, and unsaturated fatty acids such as gondoic acid,

Table 1 — Physicochemical composition of *Aesculus indica*^{63,64}

Seed elements	Value	Parameters	Seed (%)	Seeds flour
Calcium	0.08%	Moisture	51.5±1.25	10.71±1.23
Nitrogen	1.15%	Ash	1.93±0.15	3.16±0.05
Phosphorus	0.18%	Crude fibre	1.73±0.19	6.34±0.22
Potassium	0.79%	Crude fat	1.30±0.43	3.27±0.39
Sulphur	0.07%	Protein	1.39±0.5	6.78±1.19
Copper	41.2 ppm	Starch	38.18	59.59
Iron	159 ppm	Carbohydrate	42.15	69.74
Manganese	6.95 ppm	Sugar	3.97	10.15
Zinc	25.6 ppm	Energy (cal/100 g)	185.86	327.51
Oil	2.02%			

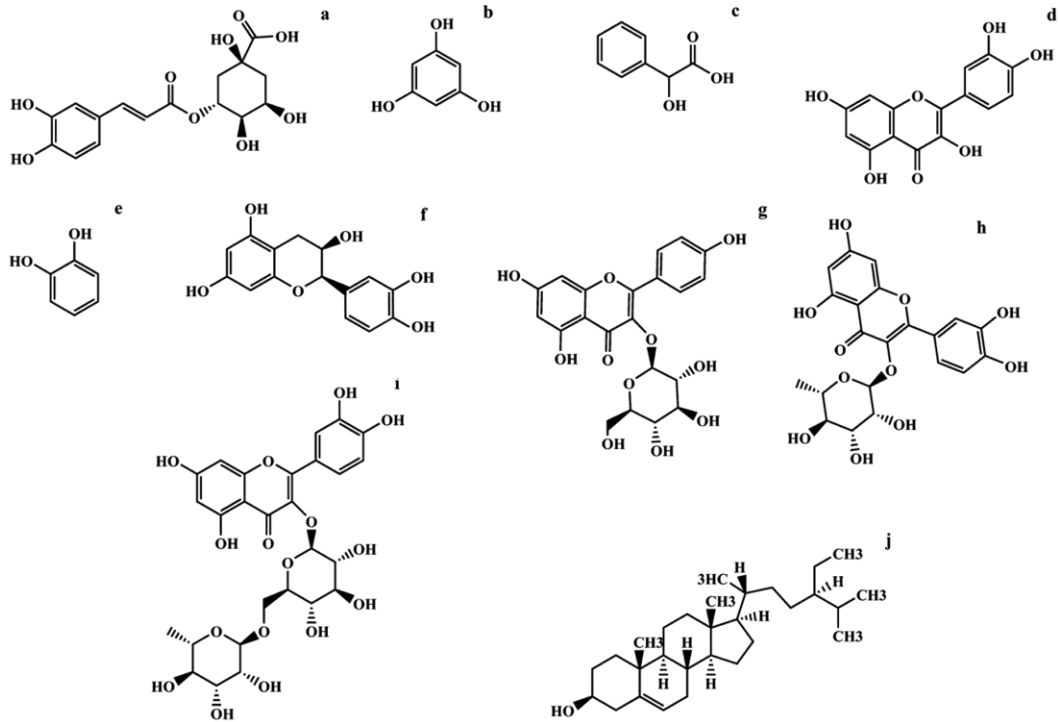


Fig. 2 — Phenol, flavonoids, and a phytosterol of *Aesculus indica*. a) Chlorogenic acid; b) Phloroglucinol; c) Mandelic acid; d) Quercetin; e) Catechol; f) Epicatechin; g) Astragalins; h) Quercitrins; i) Rutin; and j) Beta sitosterol.

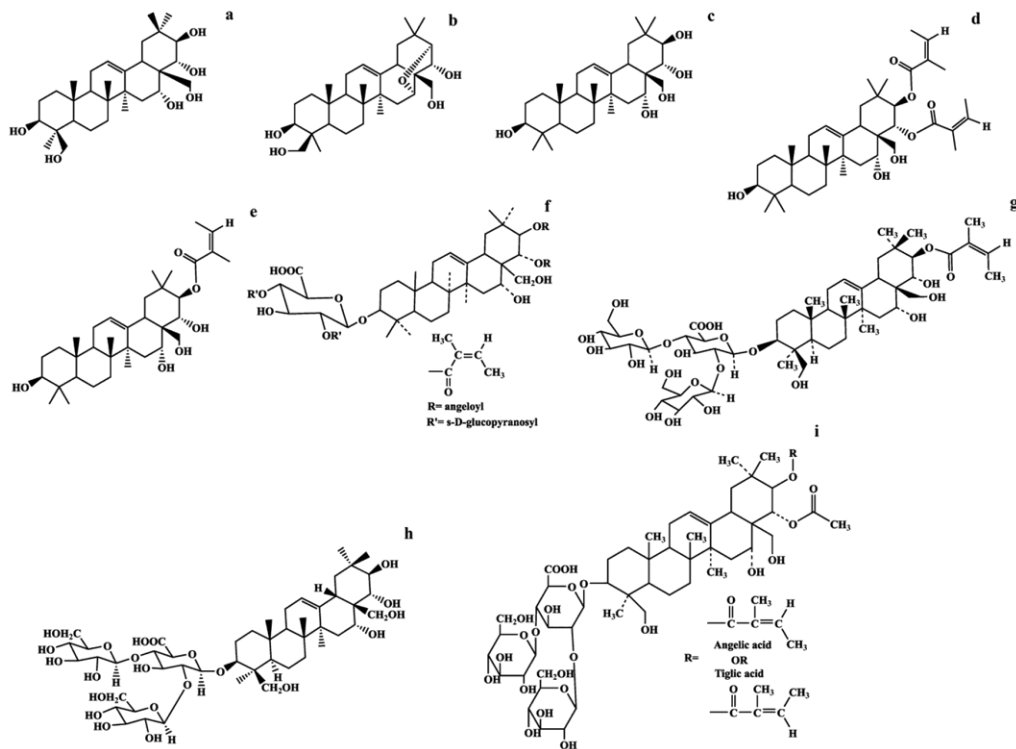


Fig. 3 — Triterpenoids and triterpene saponins of *Aesculus indica*, a) Protoaescigenin; b) Aescigenin; c) Barringtogenol-C; d) 21, 22 diangeylelbarringtogenol-C; e) 21-angeloylbarringtogenol-C; f) 3-O-[[Beta-3-D-glucopyranosyl(1-2)] {Beta-D-glucopyranosyl(1-4)}-Beta-D-gluconopyranosyl] 21, 22-diangeylelbarringtogenol C; g) Aesculusides-A; h) Aesculusides-B; and i) Aescin.

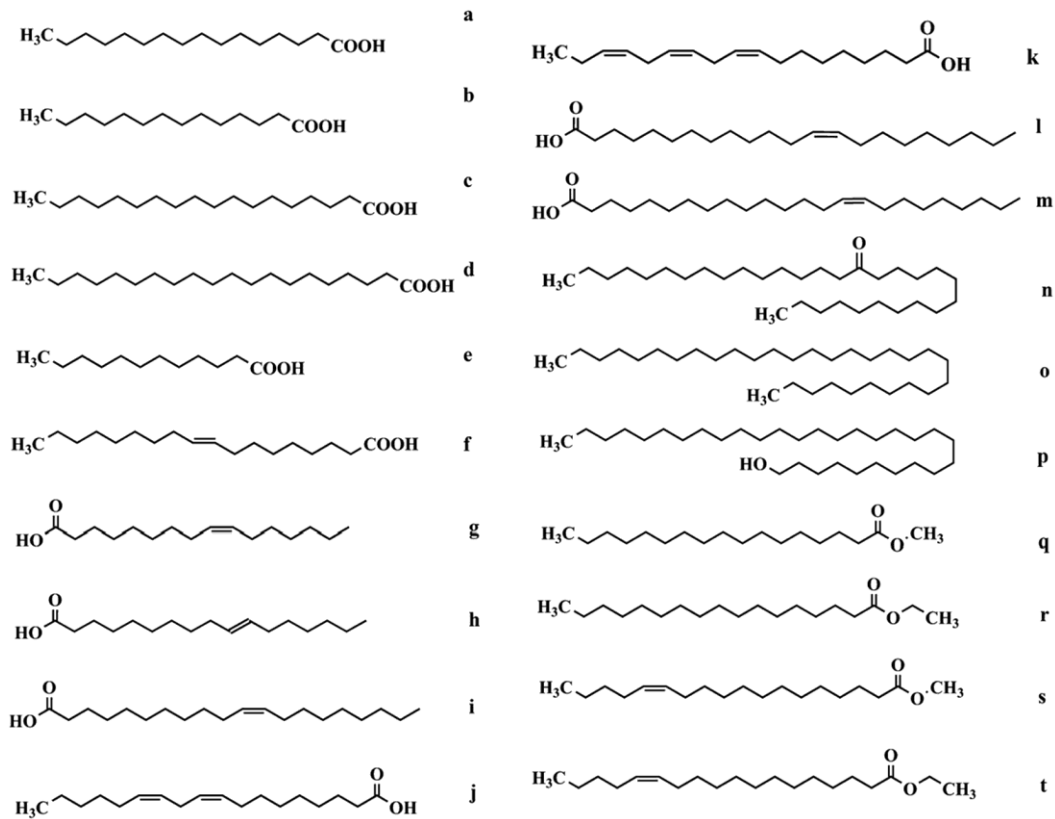


Fig. 4 — Fatty compounds of *Aesculus indica*, a) Palmitic acid; b) Myristic acid; c) Stearic acid; d) Arachidic acid; e) Lauric acid; f) Oleic acid; g) Palmitolic acid; h) 10-heptadenoic acid; i) Gondoic acid; j) Linoleic acid; k) Linolenic acid; l) Erucic acid; m) Nervonic acid; n) Palmitone; o) N-hentriacontane; p) N-hentriacontanol; q) Methyl heptadecanoate; r) Ethyl heptadecanoate; s) Methyl octadec-(13Z)-enoate; and t) Ethyl octadec-(13Z)-enoate.

linoleic acid, linolenic acid, erucic acid, nervonic acid (Fig. 4g-m) were identified, as well as their composition was determined from the petroleum seed extract³⁴. Bhattacharya *et al.*⁷⁸, found palmitone, n-hentriacontanol, n-hentriacontane (Fig. 4n-p) in the leaves, and Singh *et al.*⁷⁹, isolated aliphatic esters, methyl heptadecanoate, ethyl heptadecanoate, methyl octadec-(13Z)-enoate, and ethyl octadec-(13Z)-enoate (Fig. 4q-t) from the chloroform seeds extract.

Pharmacological properties

A. indica has exhibited various activities.

Anti-diabetic effect

Mishra *et al.*³¹, orally administrated 25, 50, and 75% processed seed flour of *A. indica* to Wistar rats and didn't find any toxic effect. However, the 75% processed seed flour diet was more effective than the 25 and 50% diet for alloxan-induced diabetic rats. There were mild swelling and necrosis in the liver, mild tubular swelling, and congestion in the kidney. The eosinophilic muscle fibres increased with the heart

inflammation, and a fully developed islet of Langerhans was observed in the pancreas. It was inferred that the activity might be due to aescin in the flour.

Enzyme inhibitory activity

The methanolic fruit extract and fractions (chloroform, ethyl acetate, aqueous, and n-hexane) of *A. indica* significantly ($P < 0.001$) inhibited the activity of acetylcholinesterase (AChE) and butyrylcholinesterase (BChE) as compared to the standard drug galanthamine. The extract showed inhibition of $IC_{50} = 145 \mu\text{g/mL}$ and $IC_{50} = 200 \mu\text{g/mL}$ and galanthamine $IC_{50} = 48 \mu\text{g/mL}$ and $IC_{50} = 66 \mu\text{g/mL}$, respectively, against AChE and BChE. However, among the fractions, the chloroform fraction displayed the highest inhibition ($IC_{50} = 85 \mu\text{g/mL}$ and $IC_{50} = 160 \mu\text{g/mL}$), followed by ethyl acetate ($IC_{50} = 190 \mu\text{g/mL}$ and $IC_{50} = 210 \mu\text{g/mL}$) and aqueous fraction ($IC_{50} = 350 \mu\text{g/mL}$ and $IC_{50} = 400 \mu\text{g/mL}$). In comparison, n-hexane exhibited the least ($IC_{50} = 760 \mu\text{g/mL}$ and $IC_{50} = 870 \mu\text{g/mL}$) inhibition, respectively, against AChE and BChE⁶⁵.

Antioxidant activity

Pal *et al.*¹⁸, reported the effect of seed (petroleum ether, chloroform, ethyl acetate, ethanol, and aqueous) extracts on (non-enzymatic haemoglobin glycosylation) antioxidant activity. The ethanol and chloroform extract exhibited the highest (76.9 ± 0.92 and 70.3 ± 0.45 respective) antioxidant activity at 1 mg/mL concentration. However, the petroleum ether, ethyl acetate, and aqueous extract showed low (45.5 ± 0.53 , 62.5 ± 0.35 , and 57.1 ± 0.40 , respectively) activity. The flavonoid and tannins detected in the extract might be responsible for antioxidant activity. In another activity, the chloroform leaf extract showed $27.62 \pm 1.67\%$ and $48.65 \pm 1.71\%$ inhibition against nitric oxide and superoxide radicals at the highest ($250 \mu\text{g/mL}$) concentration. The activity increased significantly ($P < 0.01$) with the increase in (20 - $250 \mu\text{g/mL}$) concentration. The IC_{50} value of $513.84 \mu\text{g/mL}$ and $235.4 \mu\text{g/mL}$ were respectively found in the extract as compared to standard rutin ($\text{IC}_{50} = 67.8 \mu\text{g/mL}$) and quercetin ($\text{IC}_{50} = 62.5 \mu\text{g/mL}$)⁸⁰.

Hypocholesterolemic and hypoglycaemic activity

The level of total blood cholesterol, triglycerides, low-density lipoprotein cholesterol (LDL), very low-density lipoprotein cholesterol (VLDL), and blood LDL/HDL cholesterol ratio was 386.2 ± 7.68 , 121.64 ± 2.95 , 327.16 ± 6.89 , 24.33 ± 0.59 , 9.44 ± 0.46 mg/dL respectively found in the serum of 1% cholesterol-fed control group rats. Which was significantly ($P \leq 0.05$) decreased to 247.73 ± 6.00 , 54.54 ± 4.22 , 197.41 ± 6.24 , 10.91 ± 0.85 , 5.02 ± 0.32 mg/dL in 75% processed *A. indica* seed flour fed group respectively. However, the level of high-density lipoproteins was 34.72 ± 1.88 mg/dL in the 1% fed cholesterol control, which was significantly ($P \leq 0.05$) increased to 39.41 ± 1.60 mg/dL in the 75% processed seed flour-fed group. The level of blood glucose significantly ($P \leq 0.05$) decreased from 228 mg/dL to 95 mg/dL in alloxan-induced diabetic rats fed with 75% processed flour. It was inferred that aescin might be responsible for these activities⁸¹.

Immunomodulatory activity

The oral administration (50 and 100 mg/kg/day) of methanolic leaves extract of *A. indica* to Wistar rats significantly (P , 0.001) increased the neutrophil adhesion at the highest dose compared to the control group. Although, the haemagglutinating antibody titre did not show any significant increase. However, the delayed-type hypersensitivity response to sheep red

blood cells exhibited a dose-dependent and significant (P , 0.001) increase in the paw volume. Thus, *A. indica* stimulated cell-mediated immunity without affecting humoral immunity⁸².

Anti-nociceptive activity

The 70% ethanolic leaves extract of *A. indica* dose-dependent (10, 30, 60 mg/Kg, i.p.) significantly ($P \leq 0.001$) decreased the acetic acid-induced writhing in balb/c mice as compared to the saline control group. The extract dose (100, 300, 600 mg/kg, i.p.) increased latency time in the hot plate test and reaction time in the tail immersion test, measured at 30, 60, 90, and 120 min intervals. The analgesic effect of the extract and standard drug tramadol was significant ($P \leq 0.001$) and almost similar when compared with saline control. The presence of flavonoids and tannins in the extract might be responsible for the anti-nociceptive effect possibly through the inhibition of peripheral and central nociception mechanisms⁸³.

In another study, ethanol seeds extract (100, 200, 300 mg/kg) showed significant ($P < 0.001$) anti-nociceptive activity compared to normal control in the hot plate test, tail withdrawal in the tail immersion test, and reduced number of writhing in acetic acid-induced writhing test in albino mice. The presence of saponins, flavonoids, tannins, and other phytochemicals in seeds, might be responsible for activity⁸⁴.

Anti-spasmogenic activity

The alcoholic leaves extract (400 and 800 $\mu\text{g/mL}$) of *A. indica* exhibited an anti-spasmogenic effect in the isolated guinea pig ileum. Extract acts as an antagonist of the spasmolytic effect produced due to acetylcholine⁸⁵. In another study, the crude extract dose-dependently reduced spontaneous contraction in isolated tissue of rabbit's jejunum, and the K^+ (80 mM) induced contraction showed EC_{50} values of 0.1083 mg/mL (0.04793-0.2447, 95% CI). They concluded that activity was due to the blocking of calcium channels⁸⁶.

Anti-inflammatory activity

A. indica extract (100 mg/kg) significantly (P , 0.03) reduced 1% carrageenan-induced paw oedema as compared to standard aspirin (12 mg/kg) drug. There was no significant difference found between 300 mg/kg *A. indica* extract and aspirin effect. Both imparted almost equal anti-inflammatory effects⁸⁷.

In another study, the methanol seed extract significantly (P , 0.001) increased proliferation, viability, and superoxide dismutase enzyme activity in

the human adipose-derived stem cells and decreased lactate dehydrogenase enzyme, reactive oxygen species release in the cells with the downregulation of nuclear factor kappa-light-chain-enhancer of activated B cells pathway (NF- κ B) markers tumour necrosis factor-alpha, interleukin-1,6 compared to the monosodium iodoacetate induced stress cells⁸⁸.

Antiviral activity

The composition of *A. indica* seeds containing β -aescin exhibited 90-94% inhibition against cucumber mosaic virus with an effective concentration of 1000-2000 ppm. The composition was 40, 50, 60, and 100% more effective than standard drugs zidovudine, amantadine, acyclovir, and 2-thiouracil respectively⁸⁹.

Antimicrobial activity

A. indica leaves extract was reported to possess a significant ($P < 0.05$) antibacterial effect. The methanolic leaves extract showed the highest 14 ± 0.5 and 14.5 ± 1 mm zone of inhibition against *Micrococcus luteus* and *Staphylococcus aureus* respectively. The chloroform fraction demonstrated the highest 14.5 ± 0.1 mm inhibition against *Pseudomonas pickettii* and aqueous fractions 16 ± 1 , 14 ± 0.5 , and 15 ± 0.5 mm against *Bacillus subtilis*, *Micrococcus luteus*, and *Salmonella Setubal* respectively. Although standard drug cefotaxime showed 33 ± 0 , 32 ± 0.01 , 31 ± 0.5 , 30 ± 0.4 and 30 ± 0.05 mm zone of inhibition against *Staphylococcus aureus*, *Pseudomonas pickettii*, *Bacillus subtilis*, *Micrococcus luteus*, and *Salmonella Setubal* respectively. The lower activity of other (ethyl acetate and methanol) fractions might be due to the low presence of phytochemicals⁹⁰.

Toxic activities

A. indica seeds are eaten by wild animals and humans. Though large consumption could cause vomiting, depression, diarrhoea, dilated pupils, poor coordination, paralysis, and stupor³². In a toxicity study, a single dose of seed extract showed LD₅₀ of 10.6 and 10.7 mg/g body weight, respectively, against chicks and hamsters. The administration for 2 consecutive days showed 6.5 mg/g LD₅₀ for chicks. The symptoms of fatigue, poor coordination of muscles, paralysis, coma, and death were reported⁹¹. At the same time, the study carried out by Mishra *et al.*³¹, reported that the oral administration of 25 to 75% processed seed flour was found nontoxic to Wistar rats, and a similar study on ethanol

seed extract 2000 mg/kg was also found safe to albino mice⁸⁴.

Cytotoxic/anticancer activity

The methanolic leaves extract of *A. indica* dose-dependently (10 μ g/mL to 500 μ g/mL) inhibited (34.2% to 94%) the growth of breast adenocarcinoma (MCF-7) cells in the MTT (3-(4,5-dimethylthiazol-2-yl)-2,5 diphenyltetrazolium bromide) assay. Among the fractions of methanol crude extract, they showed significant ($P \leq 0.05$) inhibition. The aqueous fraction showed the highest (59%) growth inhibition, followed by the methanol fraction (53%), ethyl acetate fraction (45%), and chloroform fraction (30.6%), while the hexane fraction showed the lowest (29.8%) inhibition. It was concluded that the high activity of the aqueous fraction might be due to the presence of saponins or flavonoids that might act on MCF-7 cells either alone or synergistically⁹². In another study, methanol seeds extract and its chloroform and ethyl acetate fractions induced cytotoxicity in the human hepatocyte carcinoma cell line (HepG2), inhibited the cell migration and colony formation, and significantly ($P < 0.001$) reduced the spheroid formation of HepG2 cells compared to the control group⁹³.

Neurodepressive activity

Piscicides are chemical substances poisonous to fish and are primarily used to eliminate the desired or unwanted fish from the pond. Bhatt⁹⁴, demonstrated that a 2.6 mg/L concentration of a piscicidal, triterpenoid glycoside of *A. indica* degraded neurons, tactile, and sensory impulses in the medulla oblongata of fish. The absence of schooling, bottom sinking, difficulty breathing, air bubbles swallowing, and slight pinkness of the gills were observed due to the damaged respiratory centre of the treated fish⁹⁴.

Insecticidal activity

Anwar *et al.*⁹⁵, reported the insect-killing effect of acidified alcoholic (ethanol + 2% acetic acid) leaf and flower extract of *A. indica*; they found that one-time topical application of 5 μ L extracts over yellow rice stem borer (*Schirpophaga incertulas*) larvae exhibited significant ($P < 0.05$) mortality 70% and 60% respectively after 48 h. Although, in the case of sugarcane mealybug (*Saccharicoccus sacchari*), where they were fed with sugarcane leaves (previously treated with extracts), the mortality of 41.67% was observed in the leaves extract after 24 h, and in the case of flower extract 75% after 12 h which

last for 24 h. The activity of flower extract was higher than the leaves extract in both insects. The presence of constituents such as aescin and saponins in the *Aesculus* species might be responsible for their surfactant property.

Discussion and future perspective

The present review highlights the information on traditional uses, nutrients, minerals, phytochemicals, and biological activities of *A. indica*. The compiled data from the literature revealed that *A. indica* has versatile uses. Especially the seed products consumed as food by folks contain various nutrients and minerals. However, seeds are also reported toxic for human consumption, so multiple methods have been used to remove toxins/bitter contents from the seeds, including traditional water wash and many other scientific methods to optimize toxicity and nutritional composition. The composition varied from method to method, so further research should be done. Similarly, other parts of *A. indica* might contain nutrients and minerals; hence, research is needed in this direction. *A. indica* exhibited anti-nociceptive, anti-inflammatory, insecticidal, antioxidant, cytotoxic, enzyme inhibitory, hypocholesterolemic, hypoglycaemic, immunomodulatory, antimicrobial, and antiviral activity, which revealed its medicinal importance. There are several research gaps found in the literature study. The mode of pharmacological action is absent in most studies, which requires further research. A comparative study with the normal cells is lacking in the cytotoxic activity of cancer cells. The enzyme inhibitory and antioxidant activities have been performed *in vitro*; therefore, further *in vivo* study is needed to prove the efficacy of extracts in the living system. In the acute oral toxicity study, 75% processed seed flour and 2000 mg/kg ethanol seed extract of *A. indica* was reported nontoxic for rodents. At the same time, biological activities such as anticancer/cytotoxic, neuro-depressive, and insecticidal showed the toxic effect of *A. indica*; thus, a detailed toxicological study is recommended. Furthermore, most of the research has been carried out on extracts or fractions, and the effect has been considered due to aescin. Undoubtedly aescin has several pharmacological properties, but there are several other phytochemicals reported in *A. indica*, so the effect might be synergic or alone or by an unknown phytochemical that cannot be neglected. Therefore, further research is required on the isolation, purification, and characterization of

compounds and their individual pharmacological and toxicological evaluation.

Conclusion

This review focuses on the traditional uses, botany, phytochemical constituents, and pharmacological attributes of *A. indica*. Literature studies have revealed that *A. indica* contains various nutrients and minerals and has exhibited significant biological activities such as enzyme inhibitory, antioxidant, immunomodulatory, hypocholesterolemic and hypoglycaemic, anti-nociceptive, anti-spasmodic, anti-inflammatory, antimicrobial, which reflects its ethnomedicinal importance. However, it has also shown neuro depressive, cytotoxic, and insecticidal toxic activities. Nevertheless, some of the traditional uses still require validation by pharmacological studies. The literature study identifies many research gaps that need further investigation.

Conflict of interest

The authors declare no conflicts of interest.

References

- 1 Khare C P, *Aesculus indica* Hook, in Indian medicinal plants an illustrated dictionary, 1st edn, (Springer, New York), 2007, 23.
- 2 Rana D, Bhatt A and Lal B, Ethnobotanical knowledge among the semi-pastoral Gujjar tribe in the high altitude (Adhwari's) of Churah subdivision, district Chamba, western Himalaya, *J Ethnobiol Ethnomed*, 2019, **15**(10), 1-21.
- 3 CABI invasive species compendium, datasheet on *Aesculus indica* (Indian horse-chestnut), Available on: <https://www.cabi.org/isc/datasheet/3317>. accessed on 22/02/2021.
- 4 Akhtar N, Rashid A, Murad W and Bergmeier E, Diversity and use of ethno-medicinal plants in the region of Swat, north Pakistan, *J Ethnobiol Ethnomed*, 2013, **9**(25), 1-13.
- 5 Zhang Z, Li S and Lian X Y, An overview of genus *Aesculus* L.: ethnobotany, phytochemistry, and pharmacological activities, *Pharm Crop*, 2010, **1**, 24-51.
- 6 Khan M P, Ahmad M, Zafar M, Sultana S, Ali M I, *et al.*, Ethnomedicinal uses of edible wild fruits (EWFs) in Swat valley, northern Pakistan, *J Ethnopharmacol*, 2015, **173**, 191-203.
- 7 Kumar R, Dey P and Agnihotri P, Glimpses on the ethnomedicinal plant diversity in Pindari valley, Uttarakhand, *Pleione*, 2020, **14**(2), 227-236.
- 8 Rawat N and Upadhaya M L, Diversity of the medicinal plants of Almora district, Uttarakhand and their ethno-medicinal use, *J Med Plants Stud*, 2020, **8**(3), 89-101.
- 9 Jamal Z, Pervez A, Hussain M, Shah G M, Shah S H, *et al.*, Ethnomedicinal plants used for gastrointestinal ailments by the rural communities of Kaghan valley, Mansehra, Pakistan, *J Appl Environ Biol Sci*, 2017, **7**(12), 41-48.
- 10 Ikram M, Screening of medicinal plants of Pakistan for anticancer activity, *Fitoterapia*, 1983, **54**, 123-126.

- 11 Singh H, Kumar A and Verma A, Evaluation of antimicrobial activity of ethanolic extract of *Aesculus indica* seeds, *World J Pharm Pharm Sci*, 2013, **2**(5), 3045-3057.
- 12 Hasija K, Lectin in agglutination and its role in detection of sugars on the surface of RBC of normal and cancer patients, *Indian J Clin Biochem*, 1991, **6**(2), 89-96.
- 13 Ikram M and Gilani S N, Anti-inflammatory activity of *Aesculus indica* fruit oil, *Fitoterapia*, 1986, **57**(6), 455-456.
- 14 Bhardwaj A, Chand P, Pakade Y B, Joshi R and Sharma M, Kinetic and equilibrium studies on adsorption of cadmium from aqueous solution using *Aesculus indica* seed shell, *Indian J Chem Technol*, 2019, **26**(2), 146-152.
- 15 Chakraborty G S, Antioxidant activity of the successive extracts of *Aesculus indica* leaves, *Int J Pharm Sci Drug Res*, 2009, **1**(2), 121-123.
- 16 Yasin G, Ahmad M and Hussain M, Pharmacological potential of plants from Himalayan region of Pakistan-assay for antioxidants indices, *Pharmacophore*, 2020, **11**(3), 82-88.
- 17 Kaur L, Joseph L and George M, Phytochemical analysis of leaf extract of *Aesculus indica*, *Int J Pharm Pharm Sci*, 2011, **3**(5), 232-234.
- 18 Pal D, Singh H and Kumar M, A preliminary study on the *in vitro* antioxidant activity of seeds of *Aesculus indica* and bark of *Populus euphratica*, *Int J Pharm Pharm Sci*, 2012, **4**(4), 249-250.
- 19 Singh H and Kumar A, Phytochemical analysis of *Aesculus indica* seeds extracts, *Innov J Ayu Sci*, 2013, **1**(1), 13-15.
- 20 Royal Botanic Gardens Kew science, plants of the world online. *Aesculus indica* (Wall. ex Cambess.) Hook, Available on: <http://www.plantsoftheworldonline.org/taxon/urn:lsid:ipni.org:names:60450399-2>. accessed on 22/02/2021.
- 21 Kumar P and Thakur N S, Studies on changes in quality characteristics of Indian horse chestnut (*Aesculus indica* Colebr.) flour during storage, *J Appl Nat Sci*, 2017, **9**(1), 445-450.
- 22 India biodiversity portal, *Aesculus indica* (Colebr. ex Cambess.) Hook, Available on: <https://indiabiodiversity.org/species/show/264509>. accessed on 22/02/2021.
- 23 Tropicos, Missouri botanical garden, Available on: <https://www.tropicos.org/name/50276950>. accessed on 22/02/2022.
- 24 Majeed M, Khan M A, Mughal A H and Bashir A, Maturity indices of Indian horse-chestnut (*Aesculus indica* Colebr) seeds under temperate Kashmir conditions, *Silva Lusitana*, 2009, **17**(2), 211-219.
- 25 Shubeena, Wani I A, Gani A, Sharma P, Wani T A, *et al.*, Effect of acetylation on the physico-chemical properties of Indian horse chestnut (*Aesculus indica* L.) starch, *Starch/Stärke*, 2015, **67**(3-4), 311-318.
- 26 Hardin J W, Studies in the Hippocastanaceae, V. Species of the old world, *Brittonia*, 1960, **12**(1), 26-38.
- 27 Parmar C and Kaushal M K, *Aesculus indica*, In *wild fruits*, (Kalyani publishers, New Delhi), 1982, 6-9.
- 28 Lone V, Tree diversity and economic importance of forest trees of Kashmir (Jammu and Kashmir), India, *Int J Fundam Appl Sci*, 2013, **2**(4), 56-63.
- 29 Mughal R, Malik A H, Dar G H and Khuroo A A, Woody flora of Poonch district in Pir Panjal Himalaya (Jammu & Kashmir), India, *Pleione*, 2017, **11**(2), 367-388.
- 30 Wani I A, Jabeen M, Geelani H, Masoodi F A, Saba I, *et al.*, Effect of gamma irradiation on physicochemical properties of Indian horse chestnut (*Aesculus indica* Colebr.) starch, *Food Hydrocoll*, 2014, **35**, 253-263.
- 31 Mishra M L, Sood S, Patial V, Sood A, Singh B, *et al.*, Histo-pathological study of alloxan-induced diabetes effect on rat's organ after intake of different proportion of *Aesculus indica* (Tatwakhar) flour, *J Entomol Zool Stud*, 2018, **6**(4), 1813-1817.
- 32 Thakur N S, Kumar P and Joshi V K, Improvement of traditional methods for the development of edible flour from Indian horse chestnut (*Aesculus indica*), *Int J Food Ferment Technol*, 2015, **5**(2), 169-176.
- 33 Rajasekaran A and Singh J, Ethnobotany of Indian horse chestnut (*Aesculus indica*) in Mandi district, Himachal Pradesh, *Indian J Tradit Knowl*, 2009, **8**(2), 285-286.
- 34 Nengroo Z R and Rauf A, Fatty acid composition and antioxidant activities of five medicinal plants from Kashmir, *Ind Crops Prod*, 2019, **140**, 111596.
- 35 Rahim I, Maselli D, Rueff H and Wiesmann U, Indigenous fodder trees can increase grazing accessibility for landless and mobile pastoralists in northern Pakistan, *Pastoralism*, 2011, **1**(2), 1-20.
- 36 Krishan R, Plants used as toxin for collection of fishes, *Int J Adv Res Sci Eng*, 2016, **5**(11), 12-24.
- 37 Negi K S and Kanwal K S, Plants used as fish toxins in Garhwal region of Uttarakhand Himalaya, *Indian J Tradit Knowl*, 2009, **8**(4), 535-538.
- 38 Matin A, Khan M A, Ashraf M and Qureshi R A, Traditional use of herbs, shrubs and trees of Shogran valley, Mansehra, Pakistan, *Pakistan J Biol Sci*, 2001, **4**(9), 1101-1107.
- 39 Rana M S, Rana S B and Samant S S, Extraction, utilization pattern and prioritization of fuel resources for conservation in Manali wildlife sanctuary, northwestern Himalaya, *J Mt Sci*, 2012, **9**, 580-588.
- 40 Sharma A, Thakur D and Uniyal S K, Plant-derived utility products: knowledge comparison across gender, age and education from a tribal landscape of western Himalaya, *J Ethnobiol Ethnomed*, 2019, **15**(1), 67.
- 41 Malik K, Ahmad M, Zafar M, Sultana S, Tariq A, *et al.*, Medicinal plants used for treatment of prevalent diseases in northern Pakistan of western Himalayas, in *Medicinal Plants - use in prevention and treatment of diseases*, (Intech Open), 2020, 1-25.
- 42 Aryal K P, Poudel S, Chaudhary R P, Chettri N, Chaudhary P, *et al.*, Diversity and use of wild and non-cultivated edible plants in the western Himalaya, *J Ethnobiol Ethnomed*, 2018, **14**(10), 1-18.
- 43 Khanday Z H and Singh S, Indigenous knowledge of medicinal plants used by tribal's and rural people of south Kashmir Himalayas, *Int J Curr Res*, 2017, **9**(8), 55469-55471.
- 44 Magar S B, Bhandari P and Ghimire S K, Ethno-medicinal survey of plants used by Magar (Kham) community, Rolpa district, western Nepal, *Ethnobot Res Appl*, 2020, **19**, 1-29.
- 45 Ahmed M J and Murtaza G, A study of medicinal plants used as ethnoveterinary: harnessing potential phytotherapy in Bheri, district Muzaffarabad (Pakistan), *J Ethnopharmacol*, 2015, **159**, 209-214.
- 46 Abbasi A M, Khan S M, Ahmad M, Khan M A, Quave C L, *et al.*, Botanical ethnoveterinary therapies in three districts of the lesser Himalayas of Pakistan, *J Ethnobiol Ethnomed*, 2013, **9**(1), 84.

- 47 Uniyal S K, Singh K N, Jamwal P and Lal B, Traditional use of medicinal plants among the tribal communities of Chhota Bhangal, western Himalaya, *J Ethnobiol Ethnomed*, 2006, **2**, 14.
- 48 Bhat J A, Kumar M and Bussmann R W, Ecological status and traditional knowledge of medicinal plants in Kedarnath wildlife sanctuary of Garhwal Himalaya, India, *J Ethnobiol Ethnomed*, 2013, **9**, 1.
- 49 Bhattarai N K, Medical ethnobotany in the Karnali zone, Nepal, *Econ Bot*, 1992, **46**(3), 257-261.
- 50 Muhammad N, Uddin N, Khan M K U, Umer M, Ali N, *et al.*, Traditional and cultural uses of medicinal plant species in the flora of Kuz Abakhel, for the treatment of various ailments, *Adv Tradit Med*, 2021, **21**, 591-607.
- 51 Muhammad N, Ali N and Uddin N, Ethno-veterinary practices used for treatment of various ailments in hilly areas of Melagah valley district Swat KPK, Pakistan, *Int J Bot Stud*, 2019, **4**(3), 171-179.
- 52 Kunwar R M, Shrestha K P and Bussmann R W, Traditional herbal medicine in far-west Nepal: a pharmacological appraisal, *J Ethnobiol Ethnomed*, 2010, **6**, 35.
- 53 Paudel H R, Poudel P, Kunwar R M, Sher H, Ur Rahman I, *et al.*, *Aesculus indica* (Wall. ex Cambess.) Hook. S apindaceae, R M Kunwar, H Sher, R W Bussmann (eds), In *ethnobotany of the Himalayas, ethnobotany of mountain regions*, (Springer, Cham), 2021, 1-9.
- 54 Shah N C and Joshi M C, An ethnobotanical study of the Kumaon region of India, *Econ Bot*, 1971, **25**(4), 414-422.
- 55 Mishra M L, Sood S, Sood A, Singh B and Shukla U N, Proximate analysis and its comparison after extraction of crude saponin/aescin different treated flour of Indian horse chestnut (*Aesculus indica*), *Int J Chem Stud*, 2019, **7**(2), 427-430.
- 56 Mishra M L, Sood S and Shukla U N, Phyto-nutritional and mineral composition of Indian horse chestnut (*Aesculus indica*) seeds, *J Pharmacogn Phytochem*, 2018, **7**(1), 2159-2162.
- 57 Mishra M L, Sood S, Sood A, Singh B, Gulati A, *et al.*, Mineral estimation of Indian horse chestnut (*Aesculus indica*) seeds after crude saponin/aescin extraction, *Int J Curr Microbiol Appl Sci*, 2018, **7**(9), 2193-2196.
- 58 Mishra M L, Sood S and Shukla U N, Standardization, development and sensory evaluation of snacks prepared from *Aesculus indica* flour (*Tatwakhar*), *Pharm Innov*, 2018, **7**(2), 89-93.
- 59 Rafiq S I, Rafiq S M and Saxena D C, Effect of hydrocolloids on the quality evaluation of flour based noodles from horse chestnut, *MATEC web conf*, 2016, **57**, 04005.
- 60 Mishra M L, Sood S and Shukla U N, Standardization, development and proximate composition of baked value added products by using Indian horse chestnut (*Aesculus indica*) flour, *Int J Curr Microbiol App Sci*, 2018, **7**(2), 1449-1458.
- 61 Shafi S, Wani I A, Gani A, Sharma P, Wani H M, *et al.*, Effect of water and ether extraction on functional and antioxidant properties of Indian horse chestnut (*Aesculus indica* Colebr) flour, *J Food Meas Charact*, 2016, **10**(2), 387-395.
- 62 Samant S S, Singh M, Lal M and Pant S, Diversity, distribution and prioritization of fodder species for conservation in Kullu District, northwestern Himalaya, India, *J Mt Sci*, 2007, **4**(3), 259-274.
- 63 Majeed M, Khan M A, Bashir A and Hussain A, Nutritional value and oil content of Indian horse-chestnut seed, *Glob J Sci Front Res*, 2010, **10**(4), 17-19.
- 64 Rafiq S I, Singh S and Saxena D C, Physical, physicochemical and anti-nutritional properties of horse chestnut (*Aesculus indica*) seed, *J Food Meas Charact*, 2016, **10**(2), 302-310.
- 65 Zahoor M, Shafiq S, Ullah H, Sadiq A and Ullah F, Isolation of quercetin and mandelic acid from *Aesculus indica* fruit and their biological activities, *BMC Biochem*, 2018, **19**(5), 1-14.
- 66 Ikram M and Khan M I, Chemical investigation of *Aesculus indica*, part II, *Fitoterapia*, 1978, **49**(6), 247-248.
- 67 Rafiq S I, Jan K, Singh S and Saxena D C, Physicochemical, pasting, rheological, thermal and morphological properties of horse chestnut starch, *J Food Sci Technol*, 2015, **52**(9), 5651-5660.
- 68 Ikram M, Khan M I and Kawano N, Chemical investigation of *Aesculus indica*, *Planta Med*, 1978, **34**(7), 337-340.
- 69 Rastogi R P and Mehrotra B N, In a compendium of medicinal plants, vol II, (CDRI, Lucknow and Publication and Information Directorate, New Delhi), 1979, 833.
- 70 Sati O P and Rana U, Triterpenoids of *Aesculus indica*, *Pharmazie*, 1987, **42**(2), 141.
- 71 Singh B, Agrawal P K and Thakur R S, Aesculuside-A, A new triterpene glycoside from *Aesculus indica*, *Planta Med*, 1986, **5**, 409-410.
- 72 Singh B, Agrawal P K and Thakur R S, Aesculuside-B, a new triterpene glycoside from *Aesculus indica*, *J Nat Prod*, 1987, **50**(5), 781-783.
- 73 Sati O P and Rana U, A new molluscicidal triterpenic glycoside from *Aesculus indica*, *Int J Crude Drug Res*, 1987, **25**(3), 158-160.
- 74 Khan L, Ahmad N, Ahmad K D, Kifayatullah Q and Arfan M, Commercial extraction of aescin, *Int J Pharmacogn*, 1995, **33**(4), 344-345.
- 75 Singh B, Simple process for obtaining beta-aescin from Indian horse chesnut (*Aesculus indica*), *US Pat 0030697*, 2006.
- 76 Costantini A, Escin in pharmaceutical oral dosage forms: quantitative densitometric HPTLC determination, *Il Farmaco*, 1999, **54**(11-12), 728-732.
- 77 Srijayanta S, Raman A and Goodwin B L, A comparative study of the constituents of *Aesculus hippocastanum* and *Aesculus indica*, *J Med Food*, 1999, **2**(2), 45-50.
- 78 Bhattacharya M K, Ghosh P K and Mukherjee K S, Chemical investigation of the leaves of *Aesculus indica*, *J Indian Chem Soc*, 1981, **58**(10), 1011-1012.
- 79 Singh B, Agrawal P K and Thakur R S, Long chain esters of *Aesculus indica*, *J Nat Prod*, 1989, **52**(1), 180-183.
- 80 Chakraborty G S, Free radical scavenging activity of *Aesculus indica* leaves, *Int J Pharm Tech Res*, 2009, **1**(3), 524-526.
- 81 Sood S, Mishra M, Sood A and Thakur V, Hypoglycaemic and hypocholesterolic efficacy of horse chestnut (*Aesculus indica*) using rat models, *J Clin Nutr Diet*, 2015, **1**(1), 1-8.
- 82 Chakraborty G S, Evaluation of immunomodulatory activity of *Aesculus indica*, *Int J Pharm Tech Res*, 2009, **1**(2), 132-134.

- 83 Firdoos S, Khan A U and Ali F, Pharmacological investigation of *Aesculus indica* aqueous-ethanol extract for its anti-nociceptive action, *J Anim Plant Sci*, 2018, **28**(2), 610-615.
- 84 More P R, Velhal A B, Chaware V J and Redasani V K, Acute toxicity study and anti-nociceptive activity of ethanol extract of *Aesculus Indica* seeds on experimental animal models, *Asian J Pharm Res Dev*, 2021, **9**(4), 31-38.
- 85 Qayum A, Ahmed N, Ahmad K D and Gilani N, Pharmacological screening of indigenous medicinal plants (III), *Pak J Pharm Sci*, 1988, **1**(1), 37-39.
- 86 Anwar A, Faisal R, Gul S, Khan F, Sultana U, *et al.*, Intestinal relaxant activity of crude extract of *Aesculus Indica* and its underlying mechanism, *Ann Punjab Med Coll*, 2022, **16**(4), 226-230.
- 87 Faisal R, Anwar A, Khokhar A, Sattar A, Jamil A, *et al.*, Anti-inflammatory activity of crude extract of *Aesculus Indica* in comparison with aspirin on carrageenan induced paw edema in rats, *Pak Armed Forces Med J*, 2022, **72**(3), 822-825.
- 88 Khawaja H, Fazal N, Yaqub F, Ahmad M R, Hanif M, *et al.*, Protective and proliferative effect of *Aesculus indica* extract on stressed human adipose stem cells via downregulation of NF- κ B pathway, *Plos One*, 2021, **16**(10), e0258762.
- 89 Singh B, Katoch M, Ram R and Aijaz Z, A new antiviral agent from Indian horse chestnut (*Aesculus indica*). European Patent Specification, *International publication no. WO 079795*, 2003.
- 90 Bibi Y, Nisa S, Chaudhary F M and Zia M, Antibacterial activity of some selected medicinal plants of Pakistan, *BMC Complement Alternat Med*, 2011, **11**, 52.
- 91 Kaushik N, Choudhary M and Jangra A, Phytochemical and pharmacological study of *Aesculus indica* linn. Plant: an updated review, *J Ayurveda*, 2021, **15**(2), 141-146.
- 92 Bibi Y, Nisa S, Zia M, Waheed A, Ahmed S, *et al.*, *In vitro* cytotoxic activity of *Aesculus indica* against breast adenocarcinoma cell line (MCF-7) and phytochemical analysis, *Pak J Pharm Sci*, 2012, **25**(1), 183-187.
- 93 Shahbaz H, Latif A, Perveen S, Hussain K, Azam S, *et al.*, *In vitro* antioxidant potential and antiproliferative activity of *Aesculus indica* seed extract against hepatocellular carcinoma cell line, *Pak J Pharm Sci*, 2021, **34**(1), 301-306.
- 94 Bhatt J P, Neurodepressive action of a piscicidal glycoside of plant, *Aesculus indica* (Colebr.) in fish, *Indian J Exp Biol*, 1992, **30**(5), 437-439.
- 95 Anwar T, Jabbar A, Khalique F, Tahir S and Shakeel M A, Plants with insecticidal activities against four major insect pests in Pakistan, *Trop Pest Manag*, 1992, **38**(4), 431-437.