

Green one-pot synthesis of cerium oxide nanoparticles utilising *Colocasia esculenta* stalk extract: An approach to antibacterial activities

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Cerium oxide (CeO₂) nanoparticles have been synthesised (NPs) via the sol-gel method. *Colocasia esculenta* (*C. esculenta*) stalk extract is employed as a green reducing and capping agent during the production. The different precursor concentrations (0.06–0.30 mol/L) are examined to prepare the NPs which exhibited an absorption band at 216 nm. Using X-ray diffraction technique, the average crystallite size of synthesised CeO₂ is found to be 6.44 nm. Transmission electron microscopy images revealed that the NPs are nearly spherical. High-intensity peaks corresponding to cerium (Ce) and oxygen (O) during energy dispersive X-ray spectroscopy assessments, confirm CeO₂ NPs production. The Fourier-transform infrared evaluations also indicated Ce-O stretching and the absence of O-N-O bending in the NPs. Furthermore, the findings demonstrated that the synthesised CeO₂ NPs exhibited susceptibility against all bacteria, except *Bacillus subtilis*.

Keywords: Antibacterial, Cerium oxide nanoparticles, Green synthesis, Stalk extract

Introduction

Research on metal oxide NPs are raising, attributable to its impressive characteristics and wide potential applications. Cerium oxide (CeO₂) is a semiconductor material with a significant energy band gap (3.19 eV) and wavelength range (330–370 nm). The material is also chemically and thermally stable^{1–3}.

Several researchers synthesised CeO₂ NPs through chemical approaches. For example, Kusuma *et al.*⁴ synthesised CeO₂ NPs with average particle size in the range 35–38 nm via sonochemical method. Those NPs were an excellent photocatalyst for wastewater treatment. Similarly, Ramachandran *et al.*⁵ reported the chemical precipitation method for synthesis of CeO₂ NPs using polyvinyl pyrrolidone as an effective capping agent. The NPs were between 19 nm and 33 nm when synthesised at pH 9 to 11. In another study, Trenque *et al.*⁶ reported the synthesis of CeO₂ NPs through pulsed laser ablation, photochemical,

and hydrothermal approaches. Novel nano-octahedra, nanocubes, or nanorods of CeO₂ NPs were synthesised. Moreover, the particles exhibited superior paraoxon degrading activities.

The effectiveness of physical and chemical techniques in producing reasonable NPs yields is hindered by their harmful precursors and notable costs and energy requirements^{7,8}. Consequently, biological or green nanomaterial synthesis procedures are gaining attention. The approaches are more cost-effective, sustainable, biocompatible, reliable, and economical than their physical and chemical counterparts. Furthermore, green synthesis techniques utilising plant extracts offer practicality, ease of handling, and cost-effectiveness without necessitating toxic substances, thus potential alternatives to conventional procedures^{9,10}.

Phytochemical-rich plant extracts could stabilise NPs during synthesis, resulting in particles with

desirable morphology and size. Plant extracts are capable of substituting chemicals and can be employed as reducing and capping agents in regulating particle formation and preventing aggregation, contributing to the stabilisation of the NPs^{11,12}. The effectiveness of plant extracts as reducing and capping agents is attributable to their significant potent phytochemical contents, such as ketones, aldehydes, flavones, amides, terpenoids, carboxylic acids, phenols, and ascorbic acids^{13,14}.

Several studies reported employing leaf extracts from *Gloriosa superba* (glory lily)¹⁵, *Acalypha indica* (Indian copperleaf)¹⁶, *Aloe vera*¹⁷, *Prosopis juliflora* Mesquite¹⁸, and *Olei europaea* olive¹⁹ for synthesis of NPs. The flower extract from *Hibiscus sabdarifa* Roselle²⁰ and leaf and flower extracts from *Calotropis procera* giant milkweed²¹ were also documented. Successful productions of nano-sized CeO₂ were also attributed to the presence of phytochemicals in the plant extracts. The compounds contain hydroxyl (OH) molecules that facilitate metal ion reductions and further regulate particle growth by capping the surfaces of the nanostructures¹.

Green CeO₂ NPs offer size and shape versatility, contributing to their adaptability for varying applications. The variations stem from several factors, such as reaction temperature and duration, solution pH, Ce salt precursor type, and phytochemical and other component concentrations extracted from the plant. Nevertheless, reports on CeO₂ NPs synthesis with *Colocasia esculenta* (*C. esculenta*) extract stalks are scarce.

C. esculenta or taro is rich in various phytochemicals, including flavonoids, alkaloids, and saponins²². The current study also adopted a simple one-pot procedure at different concentrations of the precursor. The optical properties of the NPs synthesised were determined with ultraviolet-visible (UV-Vis) spectroscopy. Furthermore, comprehensive evaluations of purity, structure, and morphology were conducted with X-ray diffraction (XRD), transmission electron microscopy (TEM), Fourier transform infrared spectroscopy (FTIR), scanning electron microscopy (SEM), and energy dispersive X-ray (EDX). The antibacterial properties of the CeO₂ NPs were also assessed against gram-negative and gram-positive bacteria. The findings could offer valuable insights into the potential applications of the nanomaterial in combating microbial infections effectively.

Experimental Section

Materials

The fresh and healthy *C. esculenta* cv Wangi stalks utilised in this study were harvested from the Malaysian Agricultural Research and Development Institute (MARDI), Serdang, Malaysia at 2°59'02.2"N 101°42'09. 97% ethanol and dimethyl sulfoxide (DMSO) were purchased from Merck, while the cerium nitrate hexahydrate (Ce(NO₃)₃·6H₂O) and acetone were purchased from Sigma-Aldrich.

Gram-positive [*Staphylococcus aureus* (*S.aureus*) and *Bacillus subtilis* (*B. subtilis*)] and Gram-negative [*Klebsiella pneumonia* (*K. pneumoniae*) and *Escherichia coli* (*E.coli*)] bacteria samples were obtained from the Culture Bank in Laboratory of Microbiology, Universiti Teknologi MARA Cawangan Negeri Sembilan, Kampus Kuala Pilah. The bacteria strains were cultivated in a Thermo Scientific™ Oxoid™ Mueller Hinton broth. All glasswares used were washed with acid followed by distilled water. Deionised (DI) water was also employed throughout the synthesis process.

Stalk extract preparation

The present study adopted the extraction procedures outlined by Mondal *et al.*²³. Firstly, the taro stalks were incised into small pieces and rinsed thrice with tap water then deionised water to remove impurities. The stalk samples were oven-dried at 80°C to remove water. Subsequently, an electric grinder was employed to ground the dried stalk into fine powder. A total of 10 g of the leaf stalk powder was transferred to a 250 mL beaker containing 100 mL distilled before being boiled at 80°C for 30 min. The dark brown *C. esculenta* leaf stalk extract solution was cooled to room temperature and filtered with Whatman filter paper. The solution was kept in a cleaned Schott bottle at 4°C before use.

Synthesis of CeO₂-NPs with *C. esculenta* stalk extract

The green and simple CeO₂ NPs synthesis approach employed in this study was based on the procedures proposed by Alam *et al.*²⁴ with minor modifications. Initially, 100 mL of the *C. esculenta* stalk extract was mixed with 0.03 M of Ce(NO₃)₃·6H₂O. The mixture was constantly agitated for 5 h at 80°C to initiate colloidal solution formation. Further continuous stirring for 30 min led to a thick yellowish-brown gel development. The mixture was left at room temperature for 2 h upon completion of

the reaction. Subsequently, the mixture was subjected to centrifugation at 3000 rpm for 2 min with ethanol and distilled water to eliminate undesired residue. Our team previously published this method²⁵; however, optimisation variables were not explored. In this study, we focus on optimising the precursor concentration to investigate its influence on the characteristics of CeO₂ NPs.

The gel was transferred to a glass Petri dish and oven-dried at 80°C overnight to ensure thorough removal of moisture. The dried precipitates were then calcined at 600°C for 2 h to enhance the crystallinity of the CeO₂ NPs yield. The yellow powder was stored in glass vials before being characterised and applied against the bacterial samples. Fig. 1 illustrates the schematic diagram of the CeO₂ NPs synthesis with the *C. esculenta* stalk extract.

Characterisation of CeO₂ NPs

The green synthesised CeO₂ NPs obtained in this study were subjected to several characterisation techniques to determine its optical properties, purity, morphology, and elemental contents. The UV-Vis spectra of CeO₂ NPs were recorded with a T80+ ultraviolet-visible (UV-Vis) spectrophotometer (PG Instruments) within the 200–400 nm range. The XRD analysis of the NPs manufactured in this study was performed using a Rigaku diffractometer. The FTIR spectra within the 500–4000 cm⁻¹ spectral range (Perkin Elmer) were used to determine the functional groups in the NPs from employing the stalk extract as the capping and reducing agents. Morphological properties and elemental compositions of the NPs were also examined with SEM and EDX (A Hitachi TM3030 PLUS model), respectively. Further morphological assessment was conducted with TEM (Talos L120C).

Antibacterial assay

The effectiveness of the green CeO₂ NPs in inhibiting bacteria was assessed qualitatively according to the method described by Dar *et al.*²⁶. An array of bacterial strains encompassing Gram-positive (*S. aureus* and *B. subtilis*) and Gram-negative species (*E. coli* and *K. pneumoniae*) were selected during the antibacterial activities evaluation. Fresh overnight 100 µL of inoculum from each bacterial sample culture was spread onto Muller Hinton agar plates. Sterile paper discs of 5 mm diameter were precisely positioned in each plate. The discs contained varying concentrations of the CeO₂ NPs (50, 100, 150, 200, and 250 µg/mL). In this study, gentamycin (10 µg) was employed as the positive control, while sterile distilled water served as the negative control. Subsequently, the plates were placed in an incubator for 18–24 h at 37±2°C. The plates were then inspected for inhibition zones around the wells. Each screening assessment was performed in triplicates.

Statistical analysis

The inhibitory zone widths and concentration values recorded in the current study were analysed with the Statistical Packages for Social Science (SPSS) software version 17 for Windows, in accordance with CLSI requirements. The results were reflected in terms of mean and standard deviation. Kruskal-Wallis evaluation was also utilised to determine differences in the sizes of the inhibitory zones produced by the CeO₂ NPs against each bacterial specimen.

Results and Discussion

Absorption spectroscopy

Fig. 2 demonstrates the absorption spectra of the green CeO₂ NPs synthesised in the present study. The maximum absorption band observed at 217 nm

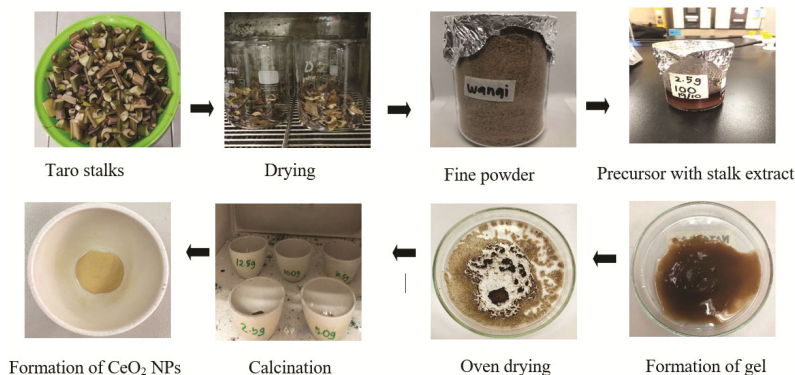


Fig. 1 — Biosynthesis of CeO₂ NPs using *C. esculenta* stalk extract

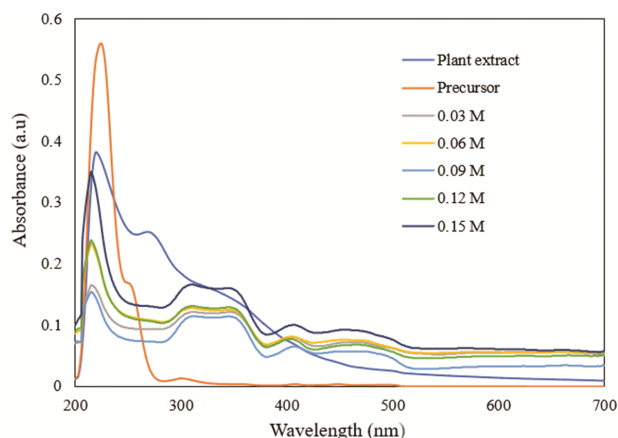


Fig. 2 — UV-visible absorption spectra of CeO₂ NPs in various concentration of precursor

Table 1 — Percentage yield (%) of the synthesised CeO₂ NPs at different concentration of precursor

Concentration of precursor (M)	Sample 1	Sample 2	Sample 3	Average
0.03	3.97	1.96	5.18	3.71
0.06	1.26	1.36	2.82	1.82
0.09	1.49	1.49	2.18	1.72
0.12	1.13	1.21	1.64	1.33
0.15	0.76	0.86	0.88	0.83

indicated the formation of CeO₂ NPs. Oxidising polyphenols, which were denoted by an expansion band, were crucial for preventing CeO₂ NPs aggregation²⁷. Similar absorption bands were recorded by several studies that reported the synthesis of CeO₂ NPs with different plants.

Sathiyapriya *et al.*²⁸ reported that CeO₂ NPs derived from *Coriandrum sativum* L. leaf extract recorded a 276 nm absorption band, indicating successful synthesis of the NPs. The results were slightly similar to earlier studies on CeO₂ NPs synthesis utilising wet chemistry²⁹, which noted the presence of CeO₂ NPs at 253 nm. Jan *et al.*³⁰ also reported a significant absorption band at 298 nm, which indicated CeO₂ NPs development. In another study, Younas *et al.*³¹ recorded a notable absorption peak (max) at 280 nm in the UV-Vis spectrum, reflecting the presence of CeO₂ NPs synthesised through the precipitation method. In the present study, the highest percentage yield was observed when the 0.03 M precursor was used (Table 1). Tesfaye *et al.*³² emphasised that factors such as precursor concentration and the ratio of plant extract significantly influenced the yield, size, shape and uniformity of NPs. Manimaran *et al.*³³ also explored optimization techniques for green CeO₂ NPs

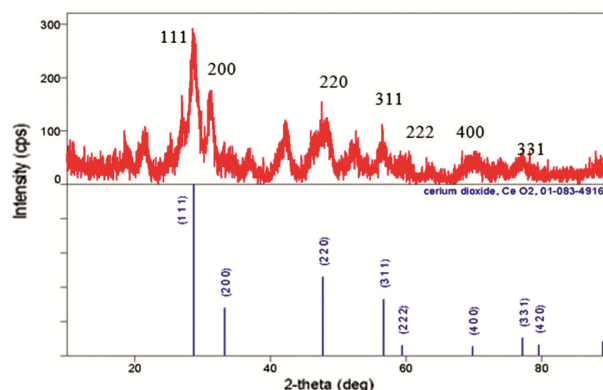


Fig. 3 — XRD pattern of CeO₂ NPs

synthesis, using a precursor concentration of 0.1, 0.2, 0.3, and 0.5 per 10 mL in *Azadirachta indica* leaf extract. In their study, the highest yield (15.43%) was obtained using optimised input parameters of 0.3 g/10 mL precursor concentration and 5 mL of *Azadirachta indica* leaf extract, centrifuged for 20 min. This highlights the critical role of precursor concentration as one of the optimization variable in the synthesis of CeO₂ NPs.

X-ray diffraction studies

Fig. 3 shows the XRD patterns of the biosynthesised CeO₂ NPs. The crystallographic arrangements of the CeO₂ NPs documented prominent diffraction peaks at 33.41°, 36.89°, 48.07°, 52.54°, 56.59°, 70.50°, and 77.23° (2θ), which corresponded to (111), (200), (220), (311), (222) (400), (331), and (420) lattice planes, respectively. The findings confirmed the crystalline nature of the CeO₂ NPs and aligned with the data reported by Zamani *et al.*³⁴. Maqbool *et al.*¹⁹ also recorded similar results for the CeO₂ NPs synthesised using *Olea europaea* leaf extract.

According to Debye-Scherrer's formula ($D=0.89\lambda/\beta \cos\theta$), the average crystallite size of the synthesised NPs were found to be 6.44 nm, which was smaller than the CeO₂ NPs produced with leaf extracts from other plants. For instance, the CeO₂ NPs synthesised utilising *Cassia angustifolia* was 36 nm³⁵. In another report, Hkiri *et al.*³⁶ noted that the CeO₂ NPs produced with *Portulaca oleracea* had crystallites of approximately 16 nm.

TEM analysis

The average size and the morphology of the synthesised CeO₂ NPs were determined through TEM analysis. Fig. 4(a) and (b) demonstrate the morphological characteristics of the biosynthesised

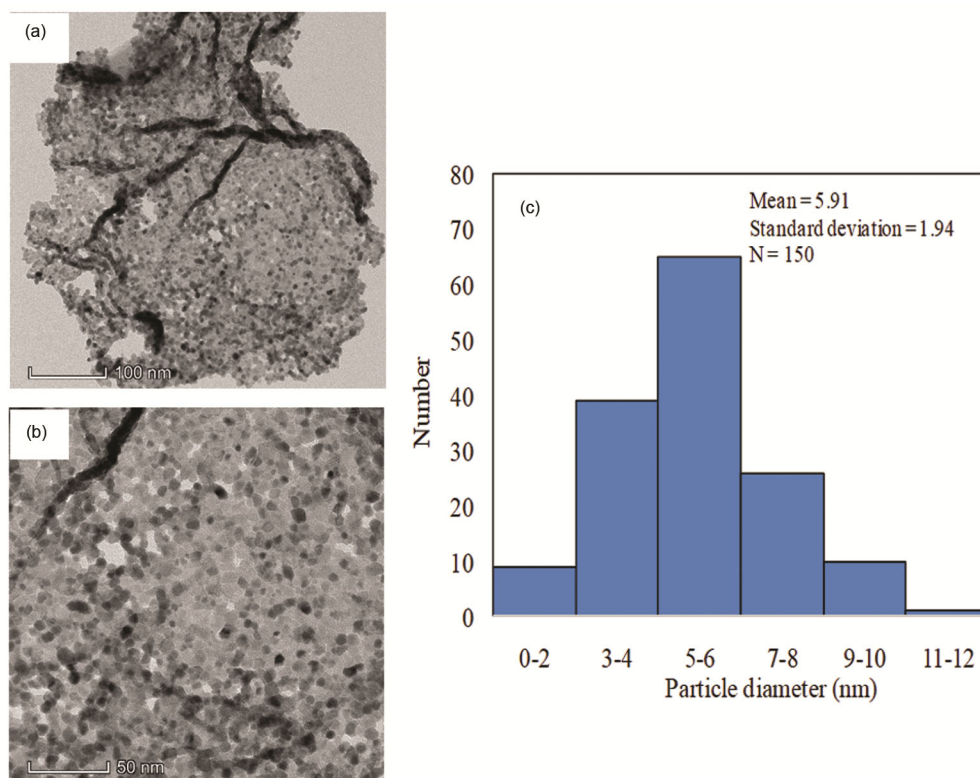


Fig. 4 — (a & b) TEM images at different magnifications and (c) corresponding histogram of CeO₂ NPs

CeO₂ NPs at different magnifications. Based on the results, most of the particles were nearly spherical. Khatami *et al.*³⁷ reported a similar spherical structure of the CeO₂ NPs synthesised utilising *Stevia rebaudiana* sugar leaf extract. Arunachalam *et al.*¹⁸ also observed spherical CeO₂ NPs derived from *Prosopis juliflora* mesquite leaf extract.

Fig. 4(c) illustrates the histogram of 150 randomly selected CeO₂ NPs distribution. The particles were analysed with ImageJ software. On average, the CeO₂ NPs was found to be 5.91 nm. The data was consistent with the figures obtained with Scherrer's formula.

FTIR analysis

The present study conducted FTIR studies to ascertain the involvement of bioactive chemicals in *C. esculenta* stalk extract in the formation of the CeO₂ NPs. The FTIR spectrum (Fig. 5) demonstrated notable absorption peaks within the 4000–500 cm⁻¹ wavenumber range. The spectral region from 2500 cm⁻¹ to approximately 4,000 cm⁻¹ corresponds to O–H and C–H stretching^{38,39}. Consequently, the extensive absorption observed within the 3750–3000 cm⁻¹ frequency range could be attributed to the stretching vibrations of O–H bonds resulting from

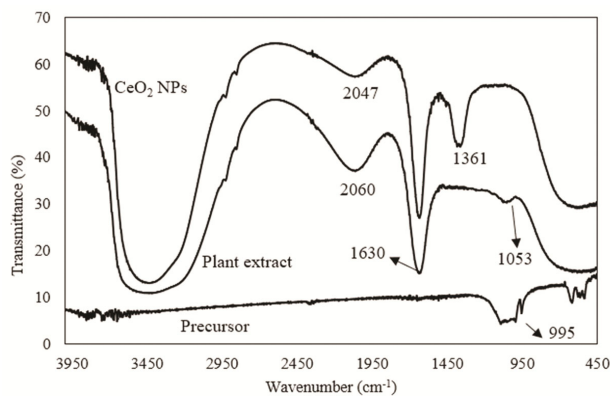


Fig. 5 — FTIR analysis of CeO₂ NPs, plant extract and precursor

residual alcohol, water, and Ce–OH groups. The functional groups in the *C. esculenta* stalk extract produced peaks at 1630 cm⁻¹ and 2047 cm⁻¹, which were associated to C=C from its phytochemical components. Similar were recorded by the CeO₂ NPs synthesised, indicating the presence of the plant extract that reduced the Ce(IV) ion to Ce metal and stabilised the resultant NPs⁴⁰. Prominent peaks observed at 3400 cm⁻¹ and 1600 cm⁻¹ also aligned with the findings reported by Kumar *et al.*⁴¹. The study utilised petals derived from the Cinnamon-like

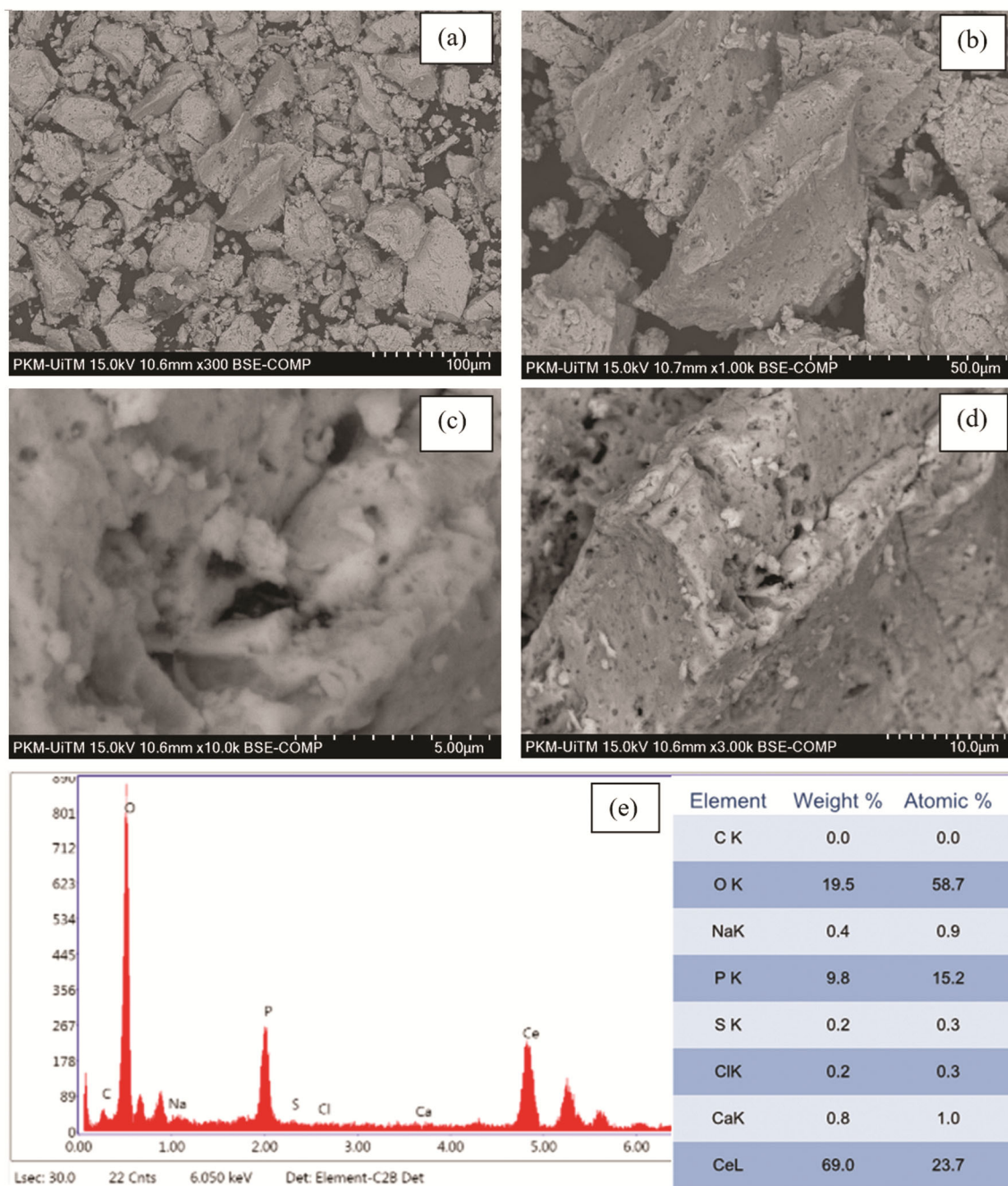


Fig. 6 — SEM of CeO₂ NPs at (a) 100 μm, (b), 50 μm, (c) 10 μm, (d) 5 μm and (e) EDX analysis

bark of *Cassia glauca* as the capping agent during synthesis of CeO₂ NPs.

SEM and EDX analysis

Fig. 6 illustrates the SEM micrographs of CeO₂ NPs at different magnifications. Based on Figs 6(a)–(d), the CeO₂ NPs had irregular shapes with slight agglomeration. The non-uniform shape of the CeO₂

NPs was comparable to the report by Alam *et al.*²⁴. Yadav *et al.*³⁵ synthesised CeO₂ NPs utilising fruit juice extract and documented agglomeration. van der Waals adhesion forces between the various non-magnetic CeO₂ NPs might led to the agglomeration⁴².

The EDX spectrum (Fig. 6e) showed high-intensity peaks of Ce (69.0%) and O (19.5%) and several impurities, such as phosphorus, calcium, sodium, and

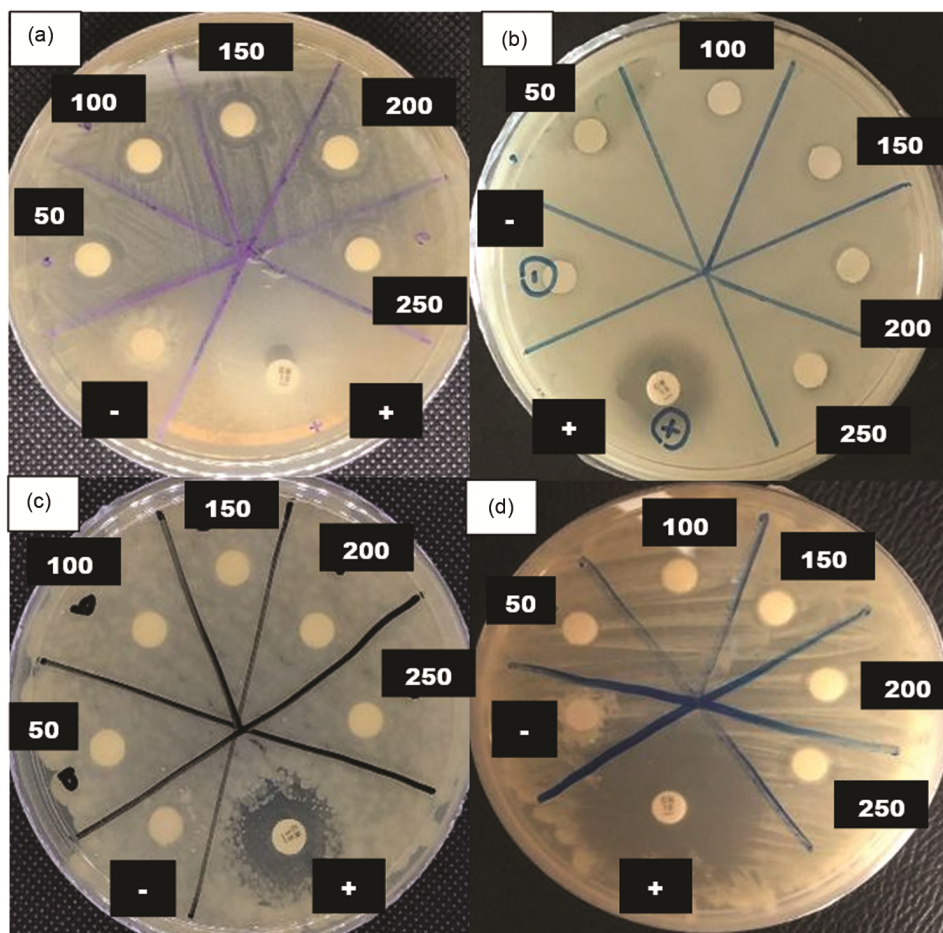


Fig. 7 — Inhibition zone of synthesized CeO₂NPs against some bacteria (a) *E.coli*, (b) *K.pneumoniae*, (c) *B.subtilis* and (d) *S.aureus*

chlorine. Farahmandjou *et al.*⁴³ also detected impurities in the CeO₂ NPs procured. The report noted Cl (0.61%), nitrogen (21.69%), P (6.57%), and potassium (6.57%) in the NPs. Thovhogi *et al.*²⁰ reported similar findings and suggested that the impurities were due to the natural organic compounds in the plant extracts employed.

Antibacterial activity

Disc-diffusion assay offers several advantages compared to other techniques, including simplicity, affordability, flexibility, and ease of data interpretation⁴⁴. Consequently, the approach is widely employed in antibacterial activity evaluations of engineered nanomaterials in laboratory settings and complex matrices⁴⁵. The antimicrobial properties of the CeO₂ NPs against Gram-positive and Gram-negative bacterial cultures were determined via the disc diffusion method and the results are shown in Fig. 7.

Although the CeO₂ NPs had adverse effect on all bacterial strains evaluated, *B. subtilis* is found to be

more resistance to the NPs. All bacterial cultures also exhibited susceptibility across CeO₂ NPs concentrations (50–250 µg/mL). Nonetheless, interactions between the NPs and the bacteria varied⁴⁶. Qi *et al.*⁴⁷ suggested that CeO₂ NPs exhibited superior efficacy against Gram-negative bacteria (*E. coli*) than Gram-positive bacteria (*B. subtilis*), as evidenced by a slight decrease in viability. The findings were also supported by the report of Sang *et al.*⁴⁸, which reported good inhibition activities of the Ce complex against *B. subtilis*.

Bacterial cell wall structures are critical in effectiveness against antibacterial activities. Gram-positive bacteria possess denser and thicker cell walls than their Gram-negative counterparts. Consequently, the microorganisms are more protected and less affected than Gram-negative bacteria by the CeO₂ NPs. Conversely, Gram-negative bacteria have thinner cell walls, hence more vulnerable to antimicrobial effects than their Gram-positive counterparts⁴⁹.

Table 2 — Antibacterial activity of biosynthesized CeO₂ NPs against pathogenic bacteria

Bacteria	Positive control (gentamicin 10 µg)	Diameter of inhibition zone (mm) of different concentration of CeO ₂ NPs (mean ± SD)					Negative control
		50 µg/mL	100 µg/mL	150 µg/mL	200 µg/mL	250 µg/mL	
<i>Klebsiella pneumoniae</i> ^{ab}	20±1.0	4.7±0.6	5.7±0.6	6.0±1.0	8.7±0.6	8.3 ±1.2	-
<i>Escherichia coli</i> ^{ab}	19±1.0	10.7±1.2	13±1.0	12.3±2.1	10.3 ±0.6	10.3±0.6	-
<i>Staphylococcus aureus</i> ^{ab}	31±5.9	3. ±1.2	3.7±0.6	7.0±1.0	6.7±1.2	13.3±0.6	-
<i>Bacillus subtilis</i> ^a	18±1.0	-	-	-	-	-	-

Note: - indicated no inhibition zone
 Values are represented as mean ± SD
 Variable with the different letter within column are significantly different at p<0.05, according to pairwise test

Ayodhya *et al.*⁵⁰ investigated the antimicrobial properties of CeO₂ NPs synthesised utilising Sapodilla fruit (*Manilkara zapota*) peel waste. The study found that *E. coli* was susceptible to the NPs, whereas *S. aureus* and *B. subtilis* were less affected due to their distinctive cell wall structure. Furthermore, electrostatic variations between the NPs manufactured in the study aided its resistance to antimicrobial treatments.

In this study, the effects of varying CeO₂ NPs concentrations (50–250 µg/mL) against the bacterial cultures were established (Table 2 and Fig. 7). The disc diffusion results documented improved inhibition zones against *E. coli* and *S. aureus* when the concentration of the NPs utilised was increased between 50 and 150 µg/mL. Roudbaneh *et al.*⁵¹ also reported similar findings.

The CeO₂ NPs at concentrations from 100 to 150 µg/mL exhibited the largest inhibition zone (10 mm) against *S. aureus*. The microorganism was also the most susceptible bacteria towards the NPs in the Gram-positive bacteria group assessed. The antibacterial activities of CeO₂ NPs against *S. aureus* were also established by Malleshappa *et al.*⁵². In the Gram-negative category, the CeO₂ NPs synthesised in this study produced the most significant inhibition zone against *E. coli* when employed at 100 µg/mL. *S. aureus* was the most affected by the CeO₂ NPs, recording maximum inhibition at 250 µg/mL of the NPs. Distinct inhibition diameters of 13 mm against *E. coli* and 13.3 mm against *S. aureus* were documented when different doses of the CeO₂ NPs were utilised. Nevertheless, no significant differences between varying CeO₂ NPs concentrations against the microorganisms were established when analysed with Kruskal-Wallis. Statistical analysis utilising Spearman's Rho indicated

non-significant correlations between the inhibition zone diameters and the bacterial strains. Furthermore, the CeO₂ NPs manufactured in the current study exhibited a weak association value across all bacteria assessed ($r = -0.150$) and inhibition zones (0.109). The negative or inverse link indicated that concentration increments lead to decreased bacterial inhibition zone diameter.

Conclusion

The current study highlighted the potential of *C. esculenta* stalk extract playing a dual-role i.e., as reducing and capping agents, during the synthesis of CeO₂ NPs. Successful NPs formation was evidenced by the absorption peak at 219 nm in the UV-visible absorption results. The average crystallite size was calculated to be 6.44 nm using Scherrer equation. XRD data aligned with the TEM images (5.91 nm), further affirming the efficacy of the synthesis method. The NPs also exhibited nearly spherical morphology, indicating precise control over particle formation with the employment of the *C. esculenta* stalk extract. Although the CeO₂ NPs demonstrated significant antibacterial activities, the nanomaterial exhibited notable resilience against *B. subtilis*. The straightforward and one-step synthesis procedure proposed in the current study offers a cost-effective, simple, safe, and appealing alternative to conventional chemical techniques.

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